spectrum was superimposable with the IR spectrum of an authentic sample. The mixed melting point obtained showed no depression.

Anal.—Calc. for $C_{15}H_{20}O_3$: C, 72.17; H, 8.06; mol. wt. 248. Found: C, 72.38; H, 8.26; m/e 248.

Isolation of Costunolide—Costunolide was isolated from the chloroform fraction of M. champaca by crystallization from hexane-benzene, and the crude crystals from T. ovata were purified by recrystallization from hexane-benzene to give 352 mg of colorless spears, mp 106–107° [lit. (4) mp 106–107°]. The IR and PMR spectra were identical to the authentic spectra.

Anal.—Calc. for C₁₅H₂₀O₂: C, 77.54; H, 8.67; mol. wt. 232. Found: C, 77.32; H, 8.84; *m/e* 232.

REFERENCES

(1) R. M. Wiedhopf, M. Young, E. Bianchi, and J. R. Cole, J. Pharm. Sci., 62, 345 (1973).

(2) T. R. Govindachari, B. S. Joshi, and V. N. Kamat, *Tetrahedron*, 21, 1509 (1965).

(3) V. Herout and F. Sorm, Chem. Ind., 1959, 1067.

(4) S. C. Bhattacharyya, G. R. Kelkar, and A. S. Rao, *ibid.*, 1959, 1069.

(5) R. I. Geran, N. H. Greenberg, M. N. MacDonald, A. M. Schumacher, and B. J. Abbott, *Cancer Chemother. Rep.*, 3 (3), 17 (1972).

ACKNOWLEDGMENTS AND ADDRESSES

Received April 2, 1976, from the Division of Pharmaceutical Chemistry, College of Pharmacy, University of Arizona, Tucson, AZ 85721. Accepted for publication July 28, 1976.

Supported by Contract NOI-CM-33750 from the Division of Cancer Treatment, National Cancer Institute, National Institutes of Health, Department of Health, Education, and Welfare, Bethesda, MD 20014, and the Elsa U. Pardee Foundation, Midland, MI 48640.

The authors are indebted to Dr. B. S. Joshi, Ciba of India Ltd., Goregaon Bombay, India, for providing the authentic sample of parthenolide, and to Professor Raymond W. Doskotch, Ohio State University, Columbus, Ohio, for providing authentic spectra (IR and NMR) of costunolide. A specimen was unavailable because of prior decomposition.

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NMR Solvent Shift Data for Methoxylated Xanthones

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Abstract \Box The NMR spectra of 1-, 2-, 3-, and 4-hydroxyxanthones, 1,3-, 1,5-, 1,6-, 1,7-, 1,8-, 2,5-, 3,4-, 3,5-, 3,6-, and 4,5-dihydroxyxanthones, 1,3,6- and 1,3,8-trihydroxyxanthones, and 1,3,6,8-tetrahydroxyxanthone, as well as those of the corresponding methyl ethers and acetates, were recorded. The spectra of the methyl ethers were measured in deuterochloroform, benzene, trifluoroacetic acid, and 3% trifluoroacetic acid in benzene. The solvent shift parameters for the methoxyl resonances are tabulated and discussed.

Keyphrases □ Methoxyxanthones, various—NMR spectra and solvent shift data □ NMR—spectra and solvent shift data, various methoxy-xanthones □ Xanthones, methoxy substituted—NMR spectra and solvent shift data

Recently, the isolation of four simple hydroxylated xanthones from seed extracts of *Mammea americana* L. was reported (1). They were identified as the 2- and 4monohydroxy derivatives and the 1,7- and 1,5-dihydroxy derivatives by a combination of spectroscopic studies and syntheses of authentic materials (1). The last substance, in fact, was prepared for the first time. The 2,5- and 4,5dihydroxy isomers were also prepared (2). During this work, other hydroxylated xanthones (3, 4) were prepared, not only for comparison purposes but also for inclusion in a preliminary antitumor screen (5).

The chemical shifts of methoxy resonances in some methoxyxanthone derivatives were dependent on the position of substitution (3). The magnitude of the benzeneinduced solvent shift for the methyl group in substituted anisoles was dependent on the nature and orientation of the substituents (6). These facts suggested that benzeneinduced shifts of methoxyl resonances in methoxyxanthones also might be position dependent and, therefore, might be helpful in elucidating the structures of naturally occurring hydroxyxanthones and methoxyxanthones.

While this work was in progress, benzene-induced shifts

Table I—Chem	uical Shifts (P	arts per Millio	on) for N	Aethoxyl
Resonances of	Various Meth	oxyxanthones	s in Vari	ous Solvents

Xanthone	Trifluoro- acetic Acid	Deutero- chloroform	Benzene	Benzene– Trifluoro- acetic Acid
1-Methoxy	4.50	4.05	3.43	3.39
2-Methoxy	4.12	3.94	3.29	3.43
3-Methoxy	4.18	3.95	3.18	3.16
4-Methoxy	4.27	4.05	3.33	3.32
1.8-Dimethoxy	4.37	3.99	3.42	3.40
3.6-Dimethoxy	4.18	3.95	3.19	3.20
4.5-Dimethoxy	4.22	4.08	3.34	3.30
1,3-Dimethoxy	4.42	4.00	3.38	3.37
, .	4.23	3.92	3.22	3.22
1,5-Dimethoxy	4.50	4.03	3.41	3.36
, .	4.28	4.03	3.34	3.30
1,6-Dimethoxy	4.44	4.03	3.45	3.45
· •	4.20	3.92	3.16	3.15
1,7-Dimethoxy	4.48	4.04	3.44	3.41
· ·	4.13	3.92	3.27	3.33
2,5-Dimethoxy	4.27	4.05	3.38	3.40
	4.12	3.95	3.29	3.33
3,4-Dimethoxy	4.30	4.05	3.84	3.75
	4.22	4.03	3.25	3.22
3,5-Dimethoxy	4.29	4.06	3.38	3.35
	4.17	3.94	3.11	3.12
1,3,6-Trimethoxy	4.38	3.99	3.44	3.42
	4.18	3.91	3.28	3.33
	4.18	3.91	3.22	3.25
1,3,8-Trimethoxy	4.34	3.98	3.43	3.45
	4.25	3.95	3.37	3.38
	4.17	3.89	3.22	3.26
1,3,6,8-Tetrameth-	4.26	3.94	3.42	3.38
oxy	4.15	3.88	3.26	3.38

of methoxyl resonances in flavonoids (7-11) and substituted xanthones (12, 13) were reported and, indeed, found to be of use in structure determination. Useful solvent shift correlations were previously derived for steroidal ketones (14, 15) and substituted coumarins (16). In addition, trifluoroacetic acid (I) was found to be a useful adjunct to the

Table II—Solvent Shift Data for Monomethoxyxanthones and Selected Dimethoxyxanthones

Xanthone	Methoxyl Position	Δ Deuterochloroform- Benzene	Δ Trifluoroacetic Acid– Benzene	۵ Trifluoroacetic Acid– Deuterochloroform
1-Methoxy	1	0.62	1.07	0.45
2-Methoxy	2	0.65	0.83	0.18
3-Methoxy	3	0.77	1.00	0.23
4-Methoxy	4	0.72	0.94	0.22
1,8-Dimethoxy	1(8)	0.57	0.95	0.38
3.6-Dimethoxy	3(6)	0.76	0.99	0.23
4,5-Dimethoxy	4(5)	0.74	0.88	0.14
1,5-Dimethoxy	1	0.62	1.09	0.47
	5	0.69	0.94	0.25
1,6-Dimethoxy	1	0.58	1.04	0.41
	6	0.76	0.99	0.28
1,7-Dimethoxy	1	0.60	1.04	0.44
	7	0.65	0.86	0.21
2,5-Dimethoxy	2	0.66	0.83	0.17
•	5	0.67	0.89	0.22
3,5-Dimethoxy	3	0.83	1.06	0.23
	5	0.68	0.91	0.23

Table III-Average Solvent Shift Values for Each Position of the Xanthone Ring

Position	Average Δ	Average ∆	Average ک
	Deuterochloroform–	Trifluoroacetic Acid–	Trifluoroacetic Acid–
	Benzene	Benzene	Deuterochloroform
1(8) 2(7) 3(6) 4(5)	$\begin{array}{c} 0.59 \; (0.57 - 0.62) \\ 0.65 \; (0.65 - 0.66) \\ 0.78 \; (0.76 - 0.83) \\ 0.71 \; (0.67 - 0.74) \end{array}$	$\begin{array}{c} 1.02 \ (0.95-1.09) \\ 0.84 \ (0.83-0.86) \\ 1.01 \ (0.99-1.06) \\ 0.91 \ (0.88-0.94) \end{array}$	$\begin{array}{c} 0.42 \ (0.38-0.47) \\ 0.20 \ (0.18-0.22) \\ 0.24 \ (0.23-0.28) \\ 0.20 \ (0.14-0.25) \end{array}$

use of benzene (II) and deuterochloroform (III) as shiftinducing solvents (9, 10, 17) and was included in the present study.

EXPERIMENTAL

The preparation of the compounds used (1-4) and their complete NMR, IR, and UV data and those of their related phenols and corresponding acetates were presented previously (4). All spectra were determined using a 60-mHz spectrometer. Tetramethylsilane (2%) was employed as the internal reference in all solvents. The concentration of the solute was not greater than 3% (w/v), and the benzene-trifluoroacetic acid solvent contained 3% (v/v) of the latter. The chemical shifts for the methyl groups in the various methoxyxanthones are listed in Table I.

RESULTS AND DISCUSSION

Table II lists the Δ values ($\Delta = \delta_{\text{Solvent 1}} - \delta_{\text{Solvent 2}}$) for the four monomethoxyxanthones and for eight (of the 10 possible) dimethoxyxanthones that do not have the methoxyl groups in the same ring. For Table III, the data from Table II were retabulated and averaged according to the methoxyl positions on the xanthone nucleus. (For the purposes of averaging, the symmetrical xanthones were counted twice. The range of values found is also included in the table.)

Since the 3(6)-position is para to the electron-withdrawing carbonyl group, it showed the largest benzene-induced shift, Δ (III–II), in line with the indications of Bowie *et al.* (6) for simple anisole derivatives. The magnitude of Δ (III–II) decreased in the order of C-3(6) > C-4(5) > C-2(7) > C-1(8), and the differences in magnitudes were large enough to distinguish among the closely related isomeric pairs listed in Table II. The introduction of the second methoxyl group into the unsubstituted ring of monomethoxyxanthone seemed to have little effect on the Δ (III–II) value for the first methoxyl. A possible exception was seen for 3,5-dimethoxyxanthone, where the shift for the 3-methoxyl group was somewhat enhanced. This exception could have been a reflection of the fact that the 5-methoxyl group, being meta to the carbonyl group, increased its electron-withdrawing power.

Methoxyl resonances appeared at relatively low field in trifluoroacetic acid, so relatively large solvent shifts were obtained in combination with benzene. These shifts again showed a dependence on the position of substitution; their magnitude decreased in the order C-1(8) \simeq C-3(6) > C-4(5) > C-2(7). The 1(8)- and 3(6)-positions (ortho and para to the

Table IV—Solvent Shift Data for S	me Methoxyxanthone	s Having More than O	ne Methoxyl Grou	p on the Same Ring
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Xanthone	Methoxyl Position	Δ Deuterochloroform- Benzene	Δ Trifluoroacetic Acid–Benzene	لے Trifluoroacetic Acid– Deuterochloroform
1,3-Dimethoxy	1	0.62	1.04	0.42
3,4-Dimethoxy	3	0.70 0.78	1.01 0.97	0.31 0.25
1,3,6-Trimethoxy	4 1 3(6)	0.21 0.55 0.63	0.46 0.94	0.19 0.39 0.97
1,3,8-Trimethoxy	6(3) 1(8) 3	0.69 0.55 0.67	0.96 0.91 0.88	0.27 0.36 0.28
1,3,6,8-Tetramethoxy	8(1) 1(8) 2(6)	0.58 0.52 0.62	0.95 0.84 0.80	0.30 0.32 0.37
Average	3(6) 3(6)	0.56	0.89	0.27
Average (12, 13)	2(7) 3(6) 4(5)	0.55 0.64 0.67	0.30	0.27

carbonyl group) were those from which the electron pairs on the methoxyl groups were most efficiently delocalized and, therefore, were the least basic positions. The fact that they showed enhanced rather than reduced values for Δ (I–II) suggests that protonation occurred on the carbonyl oxygen rather than on the ether oxygens.

Tables II and III also list the Δ (I-III) values, which also showed positional dependence, in this case mainly for the 1(8)-position where the shift was about twice that for the other three positions. This pattern was similar to that found (9) for methoxyflavones, where the shift for the 5-methoxyl (*ortho* to the carbonyl) was larger and easily distinguished from those for methoxyls at the 6-, 7-, or 8-position. Values of 0.37 and 0.15 ppm were reported (10) for the 1- and 7-methoxyls of 1,7-dimethoxyxanthone, and the deviations from the presently reported measurements of 0.44 and 0.21 ppm probably resulted from the slightly different experimental conditions.

Table IV gives shift data for a few xanthones having two methoxyls in the same ring. The abnormally low value of Δ (III-II) for the 4-methoxyl in 3,4-dimethoxyxanthone is consistent with values reported for similarly situated methoxyls flanked by two *ortho*-substituents (6, 12, 13, 16, 18). This apparent decrease in the ability of the benzene solvent to shield the methoxyl group in question has been ascribed to steric hindrance. This effect is also apparent in the corresponding value of Δ (I-II) for the 4-methoxyl in this compound. The Δ (I-III) value (0.19 ppm), however, is quite normal for a methoxyl in this position (*cf.*, the value 0.20 ppm in Table III), although it was suggested (9, 17) that such a methoxyl ought to show an enhanced shift because of its increased basicity.

Although the data in Table IV are limited, they suggest that polyoxy genation tends to increase the Δ values by a small amount. This decrease can also be noted in the values reported (12, 13) for a series of 1,3,5,6- and 1,3,6,7-tetraoxygenated xanthones (further substituted by saturated or unsaturated alkyl groups), which are included (as average values) in Table IV.

Finally, the trifluoroacetic acid addition shifts, Δ (II–II/I), were extremely small (cf., Table I), mostly within the experimental error of 1–2 Hz. Therefore, they are of little help in distinguishing the positions of methoxylation, at least in these simple xanthones. This finding is in contrast to their potential usefulness in the methoxyflavone series (9).

REFERENCES

(1) R. A. Finnegan and J. K. Patel, J. Chem. Soc. Perkin I, 1972, 1896.

(2) R. A. Finnegan and K. E. Merkel, J. Org. Chem., 37, 2986 (1972).

(3) J. K. Patel, M.S. thesis, State University of New York at Buffalo, Buffalo, N.Y., 1967.

(4) K. E. Merkel, Ph.D. thesis, State University of New York at Buffalo, Buffalo, N.Y., 1970.

(5) R. A. Finnegan, K. E. Merkel, and J. K. Patel, *J. Pharm. Sci.*, **62**, 483 (1973). *Cf.* also: R. A. Finnegan, K. E. Merkel, and N. Back, *ibid.*, **61**, 1599 (1972).

(6) J. H. Bowie, J. Ronayne, and D. H. Williams, J. Chem. Soc. B, 1966, 785.

- (7) R. G. Wilson, J. H. Bowie, and D. H. Williams, *Tetrahedron*, 24, 1407 (1968).
- (8) P. J. Garrat, F. Scheinmann, and F. Sondheimer, *ibid.*, 23, 2413 (1967).

(9) R. G. Wilson and D. H. Williams, J. Chem. Soc. C, 1968, 2477.

(10) D. Anker, C. Mercier, M. Baran-Marszak, and J. Massicot, Tetrahedron, 25, 5027 (1969).

(11) A. Pelter, R. Warren, J. N. Usmani, M. Ilyas, and W. Rahman, Tetrahedron Lett., 1969, 4259.

(12) B. Jackson, H. D. Locksley, and F. Scheinmann, J. Chem. Soc. C, 1967, 2500.

(13) F. Scheinmann, Chem. Commun., 1967, 1015.

(14) J. D. Connolly and R. McCrindle, Chem. Ind. (London), 1965, 379.

(15) D. H. Williams and N. S. Bhacca, Tetrahedron, 21, 2021 (1965).

(16) R. Grigg, J. A. Knight, and P. Roffey, ibid., 22, 3301 (1966).

(17) R. G. Wilson and D. H. Williams, J. Chem. Soc. C, 1968, 2475.

(18) H. M. Fales and K. S. Warren, J. Org. Chem., 32, 501 (1967).

ACKNOWLEDGMENTS AND ADDRESSES

Received July 1, 1976, from the Department of Medicinal Chemistry, School of Pharmacy, State University of New York at Buffalo, Buffalo, NY 14214.

Accepted for publication August 13, 1976.

Abstracted in part from a thesis submitted by K. E. Merkel to the Department of Medicinal Chemistry, State University of New York at Buffalo, in partial fulfillment of the Doctor of Philosophy degree requirements.

Supported by Grant GM-11412 from the National Institutes of Health, U.S. Public Health Service, Bethesda, MD 20014.

This article was written while R. A. Finnegan was a Guest Professor at the Institut für Pharmazeutische Arzneimittellehre der Universität München and he wishes to thank the Directors of the Institute for their hospitality.

This article is considered Part XIII in the series "Constituents of *Mammea americana* L." For Part XII, see Ref. 5.

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